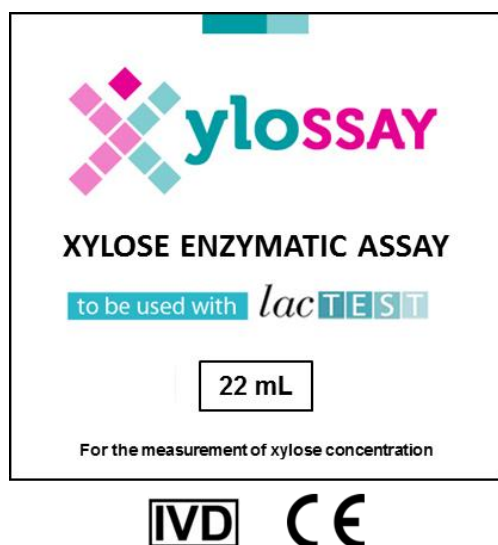


VERSION FOR ROCHE COBAS c501/c502 AUTOMATED ANALYZERS

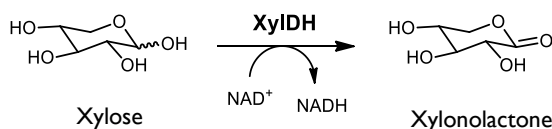


DESCRIPTION:

Xylossay[®] is a reliable and automatable method for the quantification of xylose present in urine. **Xylossay**[®] is the recommended method for xylose detection after the administration of **LacTEST 0.45 g**.

METHOD RATIONALE:

The enzyme xylose dehydrogenase (XylDH)¹ catalyzes the specific oxidation of xylose to yield xylonolactone². As XylDH is a NAD⁺ dependent enzyme, this reaction requires a concomitant reduction of this cofactor to NADH:



Production of NADH can be spectrophotometrically detected and quantified by the increase of the absorbance at 340 nm during the reaction. This way, absorbance increments will be directly proportional to the amount of xylose present in the sample.

COMPONENTS OF THE KIT:

Xylossay[®] is supplied in a format suitable for performing 70 xylose determinations. Each kit contains a vial of each one of the following components:

| VIAL | CONTENT AND FORMAT | AMOUNT |
|--------|---|---------------------------|
| Brown | Phosphate buffer, 50 mM, pH 8.0 (Solution) | 22 mL |
| Orange | β-Nicotinamide Adenine Dinucleotide (NAD ⁺) (Lyophilized) | 66.34 mg ± 12% |
| Purple | D-Xylose Dehydrogenase (Lyophilized) | 0.6 mg ± 10% |
| Pink | Calibrator: D-(+)-Xylose (Solution) | 2 mL; 3.75 mg/dL ± 5%. |

1. Sánchez-Moreno, I. et al. *J. Biotechnol.* (2016) 234:50-57.

2. Stephens, C. et al. *J. Bacteriol.* (2007) 189:2181-2185.

NON SUPPLIED ADDITIONAL EQUIPMENT:

Preparation of the reagents requires the use of micropipettes or systems suitable for dispensing the specified volumes. Vortex agitators can be used when indicated. Xylose quantification controls.

PREPARATION OF REAGENTS:

IMPORTANT: the preparation of the reagents must be conducted in the following order:

1. Dissolve the content of the purple vial (Xylose dehydrogenase, lyophilized) in **5 mL** of buffer (Phosphate buffer 50 mM, pH 8.0; brown vial). Mix gently to avoid loss of activity in the suspended enzyme. Keep cold during use if possible. This solution will be called **REACTIVE 2**.

2. Add 2 mL of buffer (Phosphate buffer 50 mM, pH 8.0; brown vial) in the vial that contains the lyophilized NAD⁺ (orange vial) and mix vigorously until its complete dissolution (vortex agitators can be used).

3. Add the totality of the 2 mL of dissolved NAD⁺ (orange vial + buffer) in the buffer vial (brown vial). Thus, the brown vial will contain **17 mL** of phosphate buffer plus NAD⁺ at the appropriated concentration. Keep cold if possible. This solution will be called **REACTIVE 1**.

4. The solution of xylose (Calibrator: standard xylose solution, pink vial) is ready for use.

CONSERVATION AND STABILITY:

Xylossay[®] must be stored at temperatures between 4° and 8°C until use, being stable under these conditions for at least 18 months. After being prepared, REACTIVES 1 and 2 are stable for at least 39 days when maintained at temperatures between 4 and 10°C (stability in automated equipments). Both REACTIVE 1 and REACTIVE 2 can be frozen at -20°C without any activity loss. Both reagents can be frozen and thawed at least 6 times without altering their function.

IMPORTANT: to use frozen reactivities, these must be completely thawed (at room temperature). Once completely thawed, they must be agitated for total homogenization before use.

ASSAY PROTOCOL:

SAMPLE PREPARATION: Collection of urine samples must be performed strictly following the specifications indicated in **LacTEST 0.45 g** summary of product characteristics. Centrifugation of urine samples is recommended (according to the protocol established in each laboratory) to remove any possible precipitate that could affect the assay. Frozen samples must be agitated after thawing for their homogenization.

PARAMETERS: wavelength 340 nm, using the standard device temperature (usually 37°C). The instrumental application for testing must be designed according to the following specifications.

CALIBRATION AND CONTROL PROCEDURES: Calibration of xylose determination in the analyzers (**CAL**) must be carried out using the standard xylose solution supplied with the kit (Calibrator, pink vial) as concentration reference (3.75 mg/dL). For calibration and xylose quantification control it is recommended the use of at least two certified and validated levels of control: a low level, with xylose concentration between 0.8 and 3 mg/dL, and a high level,

with xylose concentration between 5 and 8 mg/dL. These controls are not supplied with the kit, and must be purchased independently.

STANDARD ASSAY PROTOCOL:

The shown volumes can be increased or reduced if the final proportion between them is maintained.

| Reactive | Blank | Sample | CAL |
|---|--------|--------|--------|
| Distilled water | 30 µL | - | - |
| REACTIVE 1 | 130 µL | 130 µL | 130 µL |
| Sample | - | 30 µL | - |
| Calibrator | - | - | 30 µL |
| Mix the reaction and incubate for 5 min. (A1) | | | |
| REACTIVE 2 | 40 µL | 40 µL | 40 µL |
| Mix the reaction and incubate for 5 min. (A2) | | | |

FINAL NUMBER OF MEASUREMENTS WITH EACH KIT:

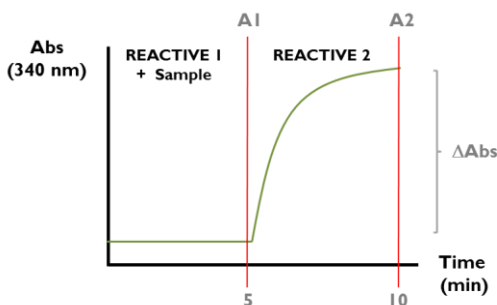
Due to the dead volume of some automated analyzers, the number of determinations can be reduced. The next formulas allow calculating the final number of assays in function of the dead volume:

$$REACTIVE\ 1 = \frac{(17\text{ mL} - \text{Dead volume in mL}) \times 1000}{\text{Volume of REACTIVE 1 in the assay } (\mu\text{L})}$$

$$REACTIVE\ 2 = \frac{(5\text{ mL} - \text{Dead volume in mL}) \times 1000}{\text{Volume of REACTIVE 2 in the assay } (\mu\text{L})}$$

CALCULATIONS:

Absorbance values (340 nm) per assay:



A1 = Initial absorbance of the mixture REACTIVE 1 + Sample (incubated for 5 min).

A2 = Final absorbance after adding REACTIVE 2 (incubated for 5 more minutes, total time of reaction: 10 min).

Differences between the two absorbance values will be proportional to xylose concentration, which can be calculated using the standard xylose solution provided with the kit (Xylose standard solution, pink vial):

$$\Delta\text{Absorbance (340 nm)} = \Delta\text{Abs} = A2 - A1$$

$$\text{Sample concentration} = [\text{Sample}] \text{ (mg/dL)}$$

$$\text{Xylose concentration (Calibrator)} = 3.75 \text{ mg/dL}$$

$$[\text{Sample}] = \frac{\Delta\text{Abs (Sample)}}{\Delta\text{Abs (Calibrator)}} \times 3.75 \text{ mg/dL}$$

The total amount of xylose in the Sample (mg) can be calculated using the total volume of urine collected during the test.

$$\text{Xylose (mg)} = [\text{Sample}] \times \text{Volume of Sample (dL)}$$

EXAMPLE:

Determination of the total amount of xylose in the urine sample of a patient:

Volume of Sample (urine) = 557 mL = 5.57 dL

- **Sample assay:** A1 = 0.122

$$A2 = 0.146$$

$$\Delta\text{Abs (Sample)} = 0.146 - 0.122 = 0.024$$

- **Calibrator assay:** A1 = 0.0870

$$A2 = 0.166$$

$$\Delta\text{Abs (Calibrator)} = 0.166 - 0.0870 = 0.079$$

$$[\text{Sample}] = \frac{0.024}{0.079} \times 3.75 \text{ mg/dL} = 1.139 \text{ mg/dL}$$

$$\text{Xylose (mg)} = 1.139 \frac{\text{mg}}{\text{dL}} \times 5.57 \text{ dL} = 6.34 \text{ mg}$$

REFERENCE VALUES

Normal values in adults:

| Test | Xylose in urine (mg) |
|----------------|----------------------|
| LacTEST 0.45 g | ≥ 19.18 |

Values below 19.18 mg indicate hypolactasia.

ANALYTICAL PARAMETERS:

The analytical parameters of **Xylossay**® application were determined during the validation of the technique in three automated analyzers (Roche Cobas c502, Werfen ILab 600 and Siemens Dimension Vista I500 analyzers).

Linearity: until at least 15 mg/dL in all the cases.

Limit of detection: between 0.13 and 0.49 mg/dL.

Reproducibility: The coefficients of variation between samples cannot be higher than 15%.

Accuracy: ≥ 88 %.

Carry-over: ≤ 4 %

ANALYTICAL PERFORMANCE:

The analytical parameters established for **Xylossay**® can slightly vary in function of the characteristics of the equipment used. Presence of some sugars as L-arabinose or glucose at high concentrations can affect xylose determination. For this reason, sample collection requires a fasting period of 10 hours. Under these conditions, no interference has been found in the collected urine.



Immunostep S.L

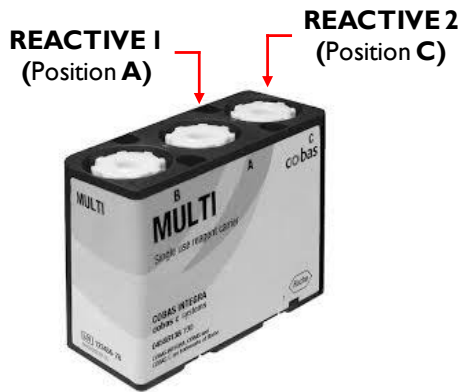
Avda. Universidad de Coimbra, s/n
Cancer Research Center (CIC)
Campus Miguel de Unamuno
37007 Salamanca (Spain)
Tel. (+34) 923 294 827
www.immunostep.com

ANNEX – APPLICATION PARAMETERS

The following parameters were optimized in a Roche Cobas c502 automated analyzer:

| | | | |
|--|------------------------|-----------------|-----------|
| Analyze | Calib. | Range | Other |
| Assay /Time /Point | 2 Point End | 10 | 32 70 0 0 |
| Wavelength (2nd/Pri.) | 415 | 340 | |
| Sample Volume | Cassette Configuration | | |
| Norm. 30.0 0.0 0 | Code 077773 | | |
| Dec. 35.0 0.0 0 | Expiration Days 99 | | |
| Inc. 30.0 30.0 0 | Reagent Volume | | |
| Dilution | R1 130 0 Inactive | Bottle Settings | |
| <input checked="" type="radio"/> Water | R2 0 0 | | |
| <input type="radio"/> Diluent | R3 40 0 | | |
| Linearity Limit | 0 % 0 % 0 0 | Version 00-01 | |
| Prozone Limit | 0 0 0 0 0 0 | Inside | 0 0 |
| Abs. Limit | 0 Increase | Save | |
| Cell Detergent | Detergent 1 | Stirring Level | 2 |
| Stirring Settings | M1 M2 M3 | | |
| UP | Stirring | Stirring | Stirring |

Cobas C Pack MULTI
(empty reagent cartridge)



| Bottle Settings | | | |
|-----------------|--------|----|------|
| Cassette Type A | | | |
| Bottle | | | |
| a | R1 | 70 | 10.0 |
| b | Cancel | 50 | 9.0 |
| c | R3 | 70 | 2.8 |
| Cancel | | OK | |

Reagent cartridge for open channel tests in Cobas equipment from ROCHE (Cobas c Pack MULTI, Roche reference: 04593138190). In the case of XYLOSSAY®, **Reagent 1** must be in **position A**, and **Reagent 2** in **position C**.

| Analyze | Calib. | Range | Other |
|---|--|--|--------------------------------|
| Calibration Type Spline | | Auto Calibration <input checked="" type="radio"/> Timeout | |
| Point | <input type="text" value="2"/> | Cassette Cancel <input type="text" value="0"/> Day | |
| Span | <input type="text" value="2"/> | Changeover Cassette Cancel | |
| Weight | <input type="text" value="0"/> | <input type="radio"/> QC Violation Method Blank Rule 1 s Control 1 None Control 2 None Control 3 None | |
| Update Type | Inside | <input type="text" value="0"/> | <input type="text" value="0"/> |
| SD Limit | <input type="text" value="999"/> | | |
| Duplicate Limit | <input type="text" value="99"/> % | <input type="text" value="32000"/> Abs. | |
| Sensitivity Limit | <input type="text" value="-99999"/> | <input type="text" value="99999"/> | |
| S1 Abs. Limit | <input type="text" value="-32000"/> | <input type="text" value="32000"/> | |
| <input type="checkbox"/> Auto Masking | | | |
| <input type="button" value="Save"/> | | | |

| Analyze | Calib. | Range | Other |
|--|--|-------------------------------------|------------------------------------|
| Application Code | XXX | | |
| Unit | mg/dl | | |
| Name | <input type="text" value="XIL-O"/> | | |
| Data mode | Active | | |
| <input checked="" type="checkbox"/> Automatic Rerun | | | |
| Technical Limit | <input type="text" value="-99999"/> | <input type="text" value="99999"/> | |
| Report Limit | <input type="text" value="-99999"/> | <input type="text" value="99999"/> | |
| <input type="checkbox"/> Control Interval Time 0 | | | |
| <input type="checkbox"/> Automatic QC On Board Stability 1 | | | |
| <input type="checkbox"/> Qualitative | | | |
| (1) 0 | L | <input type="text" value="0"/> | |
| (2) 0 | H | <input type="text" value="0"/> | |
| (3) 0 | I | <input type="text" value="0"/> | |
| (4) 0 | | | |
| (5) 0 | | | |
| (6) 0 | | | |
| Expected Values | | | |
| Male | | | |
| <input type="text" value="99"/> | Año | <input type="text" value="-99999"/> | <input type="text" value="99999"/> |
| <input type="text" value="100"/> | Año | <input type="text" value="-99999"/> | <input type="text" value="99999"/> |
| Female | | | |
| <input type="text" value="99"/> | Año | <input type="text" value="-99999"/> | <input type="text" value="99999"/> |
| <input type="text" value="100"/> | Año | <input type="text" value="-99999"/> | <input type="text" value="99999"/> |
| Default | | | |
| Sex <input type="radio"/> Male <input type="radio"/> Female | | | |
| Range <input checked="" type="radio"/> Range 1 <input type="radio"/> Range 2 <input type="radio"/> Range 3 | | | |
| <input type="button" value="Save"/> | | | |

| Analyze | Calib. | Range | Other | | | |
|---------------------------------|-------------------|-------|-------|------|------|----------------|
| Standards | | | | | | |
| Calibration Code ^[a] | XXX | XXX | 0 | 0 | 0 | 0 |
| Concentration | 0.0 | 3.75 | | | | |
| Rack No. – Pos. ^[b] | Sxxxxx-x Sxxxxx-x | | | | | |
| Sample Volume | 30.0 | 30.0 | 10.0 | 10.0 | 10.0 | 10.0 |
| Diluted S. Volume | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| Diluent Volume | 0 | 0 | 0 | 0 | 0 | 0 |
| | | | | | | Guardar |

[a] Configure in the Calibration Main Menu, Install Screen, and "Add Calibrator" window.

[b] Configure in the Calibration Main Menu, Calibrators Screen, and "Assign Calibrator" window.